Quantitative Analysis of Benzodiazepines in Whole Blood by QuEChERS and LC-MS/MS



UCT Part Numbers

ECQUUS15CT

Enviro-Clean® QuEChERS 15 mL centrifuge tube with 400 mg MgSO₄ and 100 mg NaOAc

SLDA100ID21-3UM

Selectra® DA HPLC column 100 x 2.1 mm, 3 μm

CUMPSC18CT

Enviro-Clean® dSPE 2 mL centrifuge tube with 150 mg MgSO₄, 50 mg PSA and 50 mg C18

SLDAGDC21-3UM

Selectra® DA guard column 10 x 2.0 mm, 3µm

SLGRDHLDR-HPOPT

Guard Column Holder

Summary:

Benzodiazepines (Benzos) are psychoactive drugs widely prescribed for treating anxiety, insomnia, agitation, seizures, muscle spasms, and alcohol withdrawal. Benzos are deemed safe and effective for short term use. However, frequent use of these drugs may lead to dependence and abuse. Because of this attribute, clinical, forensic and toxicological laboratories are interested in monitoring these compounds in biological samples. Common sample preparation methods for biological samples include a protein precipitation step followed by liquid-liquid extraction (LLE) or solid phase extraction (SPE). This application describes an easy, fast, and effective method using QuEChERS for the quantitative analysis of benzodiazepines in whole blood.

1 mL of negative whole blood sample is extracted using 2 mL of acetonitrile (MeCN) with 0.4 % formic acid (FA). 400 mg magnesium sulfate (MgSO₄) and 100 mg sodium acetate (NaOAc) (pre-packed in 15-mL centrifuge tube) are employed to enhance the phase separation and the partition of benzodiazepines into the organic phase. After shaking and centrifuging, 1 mL of the supernatant is purified by a 2-mL dSPE tube containing 150 mg MgSO₄, 50 mg PSA, and 50 mg C18. MgSO₄ absorbs residual water in the extract, while PSA and C18 remove organic acids and fatty matrix co-extractives, resulting in a clean extract for LC-MS/MS analysis.

Matrix matched calibration curves were constructed for the benzodiazepines quantification. The responses for 10 representative compounds were linear with R^2 ranging from 0.9963 to 1.0000 over the concentration range of 10 - 500 ng/mL. Matrix effects were evaluated by comparing the slopes of the matrix matched calibration curves to those of the calibration curves of solvent standards. The matrix effects were found to be minor, ranging from -22 to 18%. This indicated that the QuEChERS method with dSPE cleanup sufficiently removed matrix interferences that may cause significant ion suppression or enhancement. Excellent recoveries (85.5 - 105%) and relative standard deviations (RSD% \leq 10.7%) were obtained. This method was also applied to 8 real whole blood samples, no benzodiazepines were detected above the limit of quantitation of 10 ng/mL.







QuEChERS Procedure:

- 1. Add 2 mL of MeCN with 0.4% FA to 15-mL centrifuge tube containing 400 mg MgSO₄ and 100 mg NaOAc
- 2. Add internal standards
- 3. Add 1 mL whole blood
- 4. Cap and shake for 1 minute at 1000 strokes/minute using a Spex 2010 Geno-Grinder
- 5. Centrifuge for 5 minutes at 3000 g

dSPE Clean-Up:

- 1. Transfer 1 mL of supernatant to 2 mL dSPE tube
- 2. Cap and shake for 1 minute at 1000 strokes/minute using a Spex 2010 Geno-Grinder
- 3. Centrifuge for 5 minutes at 3000 g
- 4. Transfer 0.4 mL of the cleaned extract into a 2-mL auto-sampler vial, add 0.4 mL of reagent water, and vortex for 30 sec.
- 5. Samples are now ready for LC-MS/MS analysis

LC-MS/MS Parameters:

HPLC Conditions					
HPLC system	Agilent 1200 Binary Pump SL				
MS system	API 4000 QTRAP (MS/MS)				
HPLC column	UCT Selectra® DA, 100 × 2.1 mm, 3 μm (UCT p/n: SLDA100ID21-3UM)				
Guard column	UCT Selectra® DA, 10 × 2.1 mm, 3 μm (UCT p/n: SLDAGDC21-3UM)				
Guard column holder	p/n: SLGRDHLDR				
Column temperature	50 °C				
Injection volume	10 μL				
Flow rate	300 μL/min				

Mobile Phase Gradient						
Time (min)	% Mobile Phase A (0.1% Formic Acid in Water)	% Mobile Phase B (0.1% Formic Acid in MeOH)				
0.0	70	30				
0.5	70	30				
2.0	25	75				
6.5	25	75				
7.0	0	100				
9.0	0	100				
10.1	70	30				
14.0	70	30				







MRM Transitions						
Compound	Rt (min)	Q1 ion	Q3 ion 1	Q3 ion 2		
7-aminoclonazepam	7.58	286.1	222.3	250.2		
α-Hydroxy-Alprazolam	9.26	325.2	297.1	216.3		
Alprazolam	9.72	309.2	205.3	281.2		
Clonazepam	9.03	316.1	270.2	241.2		
Diazepam	9.87	285.1	193.2	154.1		
Lorazepam	8.94	321.1	303.3	275.0		
Midazolam	8.53	326.0	291.0	222.0		
Nordiazepam	9.30	271.1	140.1	165.2		
Oxazepam	9.00	287.1	241.3	104.2		
Temazepam	9.45	301.1	255.2	177.2		
Alprazolam D5	9.69	314.2	286.3	NA		
Oxazepam D5	8.98	292.1	246.2	NA		

Results:

Linearity and Matrix Effect						
Solvent standard			Matrix-matched standard			
Compound	Slope	Linearity (R²)	Slope	Linearity (R²)	Matrix Effect (%)	
7-aminoclonazepam	0.00823	0.9993	0.00646	0.9998	-22	
α-Hydroxy-Alprazolam	0.00646	0.9990	0.00764	0.9996	18	
Alprazolam	0.000413	0.9990	0.000486	0.9989	18	
Clonazepam	0.00443	0.9995	0.00497	0.9999	12	
Diazepam	0.0133	0.9997	0.0146	0.9996	10	
Lorazepam	0.00306	0.9999	0.0034	0.9997	11	
Midazolam	0.00656	0.9989	0.00675	0.9963	3	
Nordiazepam	0.00703	0.9999	0.00754	0.9998	7	
Oxazepam	0.00987	1.0000	0.0107	1.0000	8	
Temazepam	0.00641	0.9998	0.00709	0.9999	11	

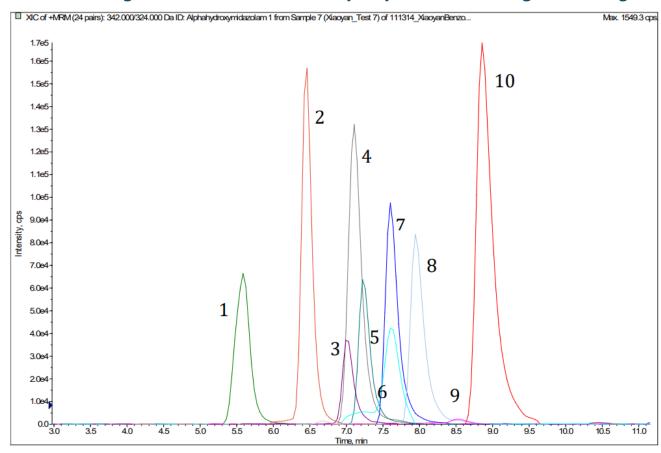
Recovery and RSD% from Whole Blood Spiked at 3 Levels (n=6)						
	10 ng/mL		50 ng/mL		200 ng/mL	
Compound	Recovery %	RSD %	Recovery %	RSD %	Recovery %	RSD %
7-aminoclonazepam	88.6	7.5	96.9	2.1	99.7	3.8
α-Hydroxy-Alprazolam	101.2	3.4	91.0	2.0	90.3	2.7
Alprazolam	92.3	10.7	90.2	4.0	86.5	3.5
Clonazepam	96.4	3.6	105.0	3.2	103.0	2.0
Diazepam	85.5	3.3	103.0	2.7	100.4	1.9
Lorazepam	96.9	5.1	93.7	4.1	91.6	2.7
Midazolam	96.7	2.7	101.6	2.7	100.6	1.9
Nordiazepam	88.4	3.9	99.7	2.5	97.8	2.3
Oxazepam	86.5	1.9	93.8	2.4	92.6	1.7
Temazepam	96.7	2.7	101.6	2.7	100.6	1.9





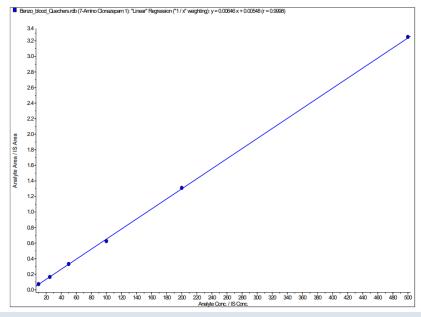


Chromatogram of a whole blood sample spiked with 200 ng/mL of drug



Peak list: 1. 7-aminoclonazepam; 2. Midazolam; 3. Lorazepam; 4. Oxazepam; 5. Clonazepam; 6. α-Hydroxy-Alprazolam; 7. Nordiazepam; 8. Temazepam; 9. Alprazolam; 10. Diazepam

Matrix Matched Calibration Curve of 7-aminoclonazepam (R²=0.9998)









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