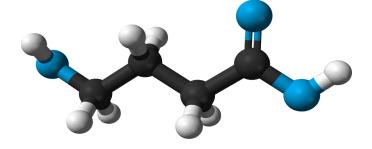
Blood GHB Extraction Procedure



UCT Part Numbers

ZSGHB020 Clean Screen® GHB 200 mg, 10 mL **SBSTFA-1-1** Selectra-Sil BSTFA with 1% TMCS 1g vial

Procedure:

1. Prepare Sample

- a) To 1 mL blood sample add internal standard and
- 0.5 mL of 100 mM phosphate buffer (pH 6.0).
- b) Mix/vortex.
- c) Rock for 10 minutes.
- d) Centrifuge for 10 minutes at 2700 rpm.

2. Condition Clean Screen® GHB Extraction Column

- a) 1 x 3 mL CH₃OH
- b) 1 x 3 mL D.I. H₂O
- c) 1 x 1 mL 100 mM phosphate buffer (pH= 6.0)
- **Note:** Aspirate at < 3 inches Hg to prevent sorbent drying.

3. Apply Sample

- a) Place centrifuge tubes into vacuum manifold for collection.
- b) The sample loading is collected.
- c) Decant sample onto column. Aspirate at about 1 inch Hg.
- d) After the sample is off the columns apply full vacuum for about 15 seconds to remove any residual blood.

4. Elute GHB

- a) Remove centrifuge tubes, set aside.
- b) Place clean centrifuge tubes into vacuum manifold for collection.
- c) 1 x 2 mL of CH₃OH/NH₄OH (99:1).
- d) Aspirate at about 1 inch of Hg.

5. Concentrate

- a) Remove test tubes from vacuum manifold.
- b) Vortex the sample prior to concentrating.
- c) Evaporate to dryness at 60 °C using a stream of nitrogen.

6. Sample Clean Up

- a) Add 200 μL of dimethylformamide.
- b) Add 1 mL of hexane saturated with dimethylformamide.
- c) Rock for 5 minutes.
- d) Centrifuge at 5 minutes at 2700 rpm.
- e) Transfer lower dimethylformamide layer to a clean test tube.

Note: If necessary transfer all liquid to a clean tube and allow to separate, then proceed to extract the lower layer

7. Concentrate

a) Evaporate to dryness at 50°C using a stream of air or nitrogen.

8. Derivatize

- a) Add 25 μL ethyl acetate and 25 μL BSTFA w 1% TMCS.
- b) Mix/vortex.
- c) Heat at 70°C for 30 minutes.

9. Quantitate

a) Inject a 1 to 2 μL of the sample onto GC/MS.





Compound	Primary Ion	Secondary	Tertiary	Cerilliant #
GHB-D6-di-TMS	239	240	241	G-006
GHB-di-TMS	233	234	235	G-001

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